Bioremediation of Wastewater Chromium through Microalgae: A Review

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Abstract - With growing industrialization and urbanization, the waste discharged to the river streams continuously pose health hazards. The wastewater discharged contains high amount of organic matter as well as heavy metals like zinc, chromium, lead, cobalt, nickel etc. Conventional methods are available to clean wastewater but they might generate large amount of sludge, leads to improper handling, disposal problem of sludge and high capital cost Therefore, there is a need of novel and sustainable technology for wastewater treatment to resolve the issue of limitations of physicochemical treatments. The use of microbial biomass (bioremediation) is considered to be a viable alternative to conventional methods.

Keywords— Pollution, Bioremediation, algae, chromium

INTRODUCTION

Environmental pollution is a very serious threat in present scenario. Rapid industrialization and urbanization are the main reason for water pollution as they are continuously discharging waste into the river. Many industries like electroplating, tanning, paper, textile etc are key component of discharging effluents causing heavy metal pollution. Heavy metal pollutants like Mercury (Hg), Cadmium (Cd), Chromium (Cr), Lead (Pb), Nickel (Ni) and Zinc (Zn) causes poisoning which can occur through drinking water or intake via food chain. These heavy metals accumulate in the food chain of aquatic and terrestrial ecosystem posing health hazards [1]. Many conventional methods are available to sequester toxic heavy metals like reverse osmosis, chemical precipitation, electro dialysis, ion-exchange and ultrafiltration. These methods and techniques are functional for removal of heavy metals with high concentration but ineffective for low concentration (1-100 ppm level) and not cost-effective. Therefore, a novel technology is needed to overcome the limitation of conventional methods which is effective and inexpensive yet providing heavy metal concentration to at least environmentally acceptable standards. Many studies have been reported investigating the potential of yeast, fungi, algae, bacteria, agricultural waste and some aquatic plants to sequester metal concentration from dilute aqueous solutions [2-4]. Out of all the heavy metals, Chromium is found to be highly toxic and carcinogenic. This paper reviews about the occurrence, sources and properties of chromium along with its immediate human health effects. The present study also gives viewpoint about how algae can be used as biosorption of chromium.

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I. CHROMIUM AS HEAVY METAL

Chromium is the 21st most abundant element in the earth's crust. It is the 24th element in the periodic table with atomic weight 52, atomic no. 24, atomic weight 51.996 g/mol and melting point 1903°C. Chromate was first discovered by L.N. Vauquelin in 1797 at Siberian red lead ore. The word chromate comes from Greek word chroma, meaning "color". Chromium is a silver grey in color, lustrous, brittle, hard metal, when heated it burns and forms the green chromic oxide. Chromium exists in oxidation state -2 to +6 but abundantly in trivalent form. Divalent chromium (+2) is unstable in most compounds as it forms trivalent compound when oxidized by air. It persists in environment in two oxidation states Cr (III) and Cr (VI) [5-6]. Hexavalent Chromium is more toxic than trivalent chromium and often present in wastewater as chromate and dichromate. Chromium is corrosion resistant. Chromium compounds have a common origin: chromite ore (Cr₂O₃, FeO). Chromium and its compounds are used in industrial sector including electroplating, chromate manufacturing, leather tanning etc [7]. In India as mentioned below in table 1, Chromate reserves are mainly (93 %) found in Odisha, mostly in the Sukinda valley in Cuttack and Jajpur districts and other reserves are located at Manipur, Nagaland, Karnataka, Jharkhand, Maharashtra, Tamil Nadu and Andhra Pradesh.

TABLE 1

Chromium compounds are found to be carcinogenic and mutagenic. Health effects of chromium compounds can vary with route of exposure. Chromium enters our body due to inhalation, ingestion or dermal contact. During occupational exposure, the absorption of inhaled Cr (VI) depends on the oxidation state, particle size, and solubility of the compound. In humans, it poses health problems like DNA damage, nausea, vomiting, nasal irritation and ulceration, skin irritation, eardrum perforation and lung carcinoma [9]. Therefore, it is very necessary to combat harmful effect of chromium which is being disposed off in water bodies through industries. In this context, Bioremediation plays an important role in sequestering chromium from wastewater. It is an innovative technology which uses the metabolic potential of microorganisms to remove toxic metals, in order to decontaminate the polluted areas [10,11]. The major

advantages of bioremediation over conventional treatment methods include low cost; high efficiency; minimisation of chemical and/ or biological sludge; no additional nutrient requirement; regeneration of biosorbent; and possibility of metal [12].

II. BIOREMEDIATION OF CHROMIUM THROUGH ALGAE

Many Studies have been carried out on application of algae like biofuel production, biomass production and assessment of water contamination with heavy metals and pesticides. Of all the microbes, algae are able to take up, accumulate and concentrate heavy metals in significant amounts from the aqueous solution. Algae have also been considered to be potential biosorbents because of their easy handling, cheap availability, relatively high surface area and high binding affinity [13,14,1]. Microalgae remove heavy metals directly from polluted water by two major mechanisms; the first is a metabolism dependent uptake into their cells at low concentrations, the second is biosorption which is a non-active adsorption process [15,16,17]. Phycoremediation is defined as a use of microalgae or macroalgae to detoxify heavy metals [18,11]. Table 2 describes that algae have many unique characteristics which make it suitable for removal of selected metals from wastewater

TABLE 2

Algae have high metal binding capacities, since polysaccharides, proteins or lipids on the surface of their cell walls have some functional groups such as amino, hydroxyl, carboxyl and sulphate, which can act as binding sites for metals The presence of functional groups with binding abilities do not always gurantee biosorption due to steric or conformational hindering or other barriers. [19,20,21,22, 23]. Algae also has the the ability to grow both autotropically *and* heterotrophically [24].

It is clear from the table above that work has been done on Chromium removal by algae e.g immobilized algal biomass was characterized for removal of chromium which resulted in maximum metal uptake of 11.494 mg/g [27]. Chlorella Pyrenoidosa was tested for its Chromium (VI) removal capacity from synthetic wastewater using immobilization (calcium alginate and carrageenan). Maximum metal uptake by the algae immobilized with calcium alginates beads was observed at pH 3 and concentration of 75mg/l [28]. Chromium removal capacity was estimated for dried green alga Ulva lactuca and activated carbon. The maximum efficiency was found to be 92% with absorption capacity of 10.61 mg/g [29]. Metal uptake studies have been conducted mainly employing laboratory grown algal species, using single metal ions. There are limited studies which employed algal species, naturally growing in polluted water to remove multimetal ions from solution. Also, there are not many studies done comparing the capacity of freshwater algae and brackish water algae to sequester heavy metal. Since industrial effluents may contain

more than one metal ion, and the algae growing in metal polluted water may have higher biosorption potential.

III. HEAVY METAL ACCUMULATION AND ITS MECHANISM

Heavy metals enter micro algal cells through micronutrient transporters. As they enter the cell, binding to specific intracellular compounds and/or transport of the metals to specific cellular compartments takes place which helps in heavy metal detoxification. Another method is chelation in which it converts heavy metals into other forms to reduce their toxicity. The detoxification mechanisms of heavy metals by algae include systems such as Metallothioneins (MTs) and Phytochelatins, Cell wall components (Alginates and guluronic acid, sulfated polysaccharides and alginates [29, 30]. The biosorption process takes place in two steps: first rapid physical adsorption (between metal ions and cell surface) and then slow chemical adsorption [31.32, 17]. The principle mechanism of removal of metallic ions involves formation of complexes between metal ions and functional groups (hydroxyl, phosphoryl, amino, carboxyl, sulphydryl, amine, imidazole, sulphate, phosphate, carbohydrate) which are present on surface of algal cell wall [33,12]. The algae and metal ion interaction depends on various factors such as physiological condition of algal cells, form of metal (chemical speciation), influence of other ions (Na²⁺, Ca²⁺), influence of trace metals, inherent tolerance of algal cells. Metal removal by biosorbents from wastewater is strongly influenced by number of abiotic and biotic factors such as pH, Chelating agents, redox potential, temperature, light, cellular activity, algal biomass concentration and extracellular products [34,35].

IV. ROLE OF MOLECULAR STUDY IN BIOREMEDIATION

Rapid Identification of specific microorganism with high affinity to absorb/ adsorb the heavy metals as well as its modification for the specific role like bioremediation is required for its commercial use. Degradation of the contaminants from the environment become much easier by using molecular ecological techniques (Fig. 1) such as direct DNA isolation from environmental samples, denaturing gradient gel electrophoresis, PCR methods and nucleic acid hybridization. These techniques have proved to have advantage over conventional methods of isolation and identification of microorganisms (taxonomical and morphological). In algal research, molecular biology acted revolutionary in studying the DNA regions by amplifying it and using Polymerase Chain Reaction (PCR). The PCR has a property of amplifying a target sequence from crude DNA model. In the laboratory, DNA fragments are synthesized and product contains many copies of fragment which can be used for identification [36,37]. Due to the limitations of morphological identification of microalgae, molecular markers such as rbcL [38] and 18s rDNA [39] has proved to be very useful. Recently, colony PCR for

microalgae has been reported using Chelex-100 for DNA extraction and amplification using the cells from the liquid cultures (Wan et al. 2011) [41].

Wu et al. 2013 [43] isolated 4 strains of Chlorella and analyzed nuclear and chloroplast encoded rDNA sequences using PCR technique. Another study was done by Ponnusawamy et al. 2013 [44] for identification of microalgae using genomic DNA, and 16S rRNA gene amplification and resultant showed Chlorella vulgaris. The 16S rRNA gene sequencing is mainly used to determine the phylogenetic of the unknown microbe among position known microorganisms. This is achieved by submitting the unknown microbe sequence to the GenBank 16S rRNA sequences database in National Center for Biotechnology Information [NCBI]. The pair of sequences from different organisms are aligned and differences in their nucleotide sequence is counted. The similarity between a pair 16S rRNA sequence is done by a comparative tool named BLAST [45,46,47]. PCR method is mainly used for tracking genetically modified microorganisms, monitoring indicator pollutants in water, soils and sediments, cloning genes, measuring gene expression by viable m/o as well as detecting specific population based on gene sequence [48,49,50]. The 18S rRNA gene sequence was studied for identification of microalgae and amplified gene sequence found to have >99% identity with Coelastrum sp. strains in the NCBI database [51].

V. CONCLUSION

Various contaminants like metals and heavy metals are constantly released into the environment by anthropogenic activities. From this exhaustive assessment of literature, it is concluded that heavy metal pollution is one of the major threat to water bodies with special reference to chromium, which is been discharged into the river streams industries. Bioremediation of heavy metals is considered to be economically promising alternative to conventional methods of remediation. There is a tremendous capacity of different species of algae in removal of chromium from wastewater which should be exploited. Molecular tools are among some steps towards phycoremediation

REFERENCES

- S.S. Ahluwalia and D. Goyal, "Microbial and plant derived biomass for removal of heavy metals from waste water", Bioresouce Technology, Vol. 98, pp. 2243-2257, 2007.
- [2] A. Mishra and A. Malik, "Recent Advances in Microbial Metal Bioaccumulation", Critical Reviews in Environmental Science and Technology, Vol. 43(11), pp. 1162-1222, 2013.
- [3] Agency for Toxic Substances and Disease Registry (ATSDR). Toxicological Profile for Chromium, U.S. Department of Health and Human Services, Public Health Service, ATSDR, September 2000.
- [4] N. Ahalya, T.V. Ramachandra and R.D. Kanamadi, "Biosorption of heavy metals", Research Journal of Chemistry and Environment, Vol. 7, pp. 71-78, 2003.
- [5] J. Barnhart, "Occurrences, Uses, and Properties of Chromium", Regulatory toxicology and pharmacology, Vol. 26, pp. S3-S7, 1997.
- [6] A.K. Shanker, C. Cervantes, L.H. Tavera and S. Avudainayagam, "Chromium toxicity in plants", Environment International, Vol. 31, pp. 739–753, 2005.
- [7] F. Baruthio, "Toxic effects of chromium and its compounds", Biological trace element research, Vol. 32, pp. 145-153, 1992.

- [8] Indian Minerals Yearbook (part II). 50th edition Chromite. Government of India, Ministry of Mines, Indian Bureau of Mines. 2011.
- [9] A.D. Dayan and A.J. Paine, "Mechanisms of chromium toxicity, carcinogenicity and allergenicity: Review of the literature from 1985 to 2000." Human & Experimental Toxicology, Vol. 20, pp. 439 – 451, 2001.
- [10] R. Kumar and D. Goyal, "Comparative biosorption of Pb²⁺ by live algal consortium and immobilized dead biomass from aqueous solution", Indian Journal of Experimental Biology, Vol. 46, pp. 690-694, 2008.
- [11] V. Sivasubramanian, V.V. Subramanian and M. Muthukumaran, "Phycoremediation of effluent from a soft drink manufacturing industry with a special emphasis on nutrient removal – a laboratory study", Journal of Algal Biomass Utilization, Vol. 3(3), pp. 21-29, 2012.
 [12] B. Volesky and Z.R. Holan, "Biosorption of heavy metals",
- [12] B. Volesky and Z.R. Holan, "Biosorption of heavy metals", Biotechnology Progress, Vol. 11, pp. 235-250, 1995.
- [13] V.J. Dominic, S. Murali and M.C. Nisha, "Phycoremediation efficiency of three micro algae *Chlorella vulgaris, Synechocystis salina* and *Gloeocapsa gelatinosa*", SB Academic Review, Vol. 16(1,2), pp. 138-146, 2009.
- [14] H. Doshi, C. Seth, A. Ray and I.L. Kothari, "Bioaccumulation of heavy metals by green algae", Current Microbiology, Vol. 56, pp. 246-255, 2008.
- [15] V.K.Gupta, A.K. Shrivastava and N. Jain, "Biosorption of chromium(VI) from aqueous solutions by green algae *Spirogyra* species", Water Research, Vol. 35 (17), pp. 4079–4085, 2001.
- [16] A.A. Hamdy, "Biosorption of heavy metals by marine algae", Current Microbiology, Vol. 41, pp. 232-238, 2000.
 [17] S.K. Mehta and J.P. Gaur, "Use of algae for removing heavy metal ions
- [17] S.K. Mehta and J.P. Gaur, "Use of algae for removing heavy metal ions from wastewater: Progress and prospects", Critical Reviews in Biotechnology, Vol. 25, pp. 113-152, 2005.
 [18] P.H.Rao, R.R. Kumar, V.V. Subramanian and V. Sivasubramanian,
- [18] P.H.Rao, R.R. Kumar, V.V. Subramanian and V. Sivasubramanian, "Environmental impact assessment of *Chlorella vulgaris* employed in phycoremediation of effluent from a leather-processing chemical industry", Journal of Algal Biomass Utilization, Vo. 1(2), pp. 42-50, 2010.
- [19] S.M. Macfie and P.M. Welbourn, "The Cell Wall as a Barrier to Uptake of Metal Ions in the Unicellular Green Alga *Chlamydomonas reinhardtii* (Chlorophyceae)", Archives of Environmental Contamination and Toxicology, Vol. 39, pp. 413-419, 2000.
- [20] K. Dayana, Ch.V. Sowjanya and Ch.V. Ramachandramurthy, "Ecofriendly remediation of industrial effluents via biosorption technology -An overview ", International Journal of Engineering Research & Technology (IJERT), Vol. 2(11), pp. 1275-1284, 2013.
- [21] V.K. Gupta and A. Rastogi, "Biosorption of lead(II) from aqueous solutions by non-living algal biomass *Oedogonium sp.* and *Nostoc sp.*: a comparative study", Colloids and surfaces B : Biointerfaces, Vol. 64, pp. 170-178, 2008.
- [22] N. Koutahzadeh, E. Daneshvar, M. Kousha, M.S. Sohrabi, and A. Bhatnagar, "Biosorption of hexavalent chromium from aqueous solution by six brown macroalgae", Desalination and Water Treatment. 2013, DOI:10.1080/19443994.2013.764353
- [23] K. Pakshirajan, A.N. Worku, M.A. Acheampong, H.J. Lubberding and N.P.L. Lens, "Cr(III) and Cr(VI) Removal from Aqueous Solutions by Cheaply Available Fruit Waste and Algal Biomass", Applied Biochemistry and Biotechnology, 2013, DOI 10.1007/s12010-013-0202-6.
- [24] K.B. Chekroun and M. Baghour, "The role of algae in phytoremediation of heavy metals: A review", Journal of Materials and Environmental Science, Vol. 4(6), pp. 873-880, 2013.
- [25] A. Sari and M. Tuzen, "Biosorption of total chromium from aqueous solution by red algae (*Ceramium virgatum*): Equilibrium, kinetic and thermodynamic studies", Journal of Hazardous Materials, Vol. 160, pp. 349–355, 2008.
- [26] A, Yaqub, M.S. Mughal, A. Adnan, W.A. Khan and K.M. Anjum, "Biosorption of hexavalent chromium by *Spirogyra* spp.: equilibrium, kinetics and thermodynamics", The Journal of animal & plant sciences, Vol. 22(2), pp. 408-415, 2012.
- [27] Kurniasih, H.D. Ariesyady, A. Sulaeman and E, Kardena, "Biosorption of Chromium (VI) Using Immobilized Algal-bloom Biomass: Kinetics and Equilibrium Studies", International Journal of Environment and Resource (IJER), Vol. 2(1), pp. 24-31, 2013.
- [28] P. Rao, V. Saisha and S.S Bhavikatti, , "Removal of Chromium (VI) from Synthetic Waste water using Immobilized Algae", International Journal of Current Engineering and Technology, 2013.

- [29] A. El-Sikaily, A. El Nemr, A. Khaled and Ola Abdelwehab, "Removal of toxic chromium from wastewater using green alga *Ulva lactuca* and its activated carbon", Journal of Hazardous Materials, Vol. 148 (1-2), pp. 216–228, 2007.
- [30] B. Volesky, "Detoxification of metal-bearing effluents: biosorption for the next century", Hydrometallurgy, Vol. 59, pp. 203–216, 2001.
- [31] P.X. Sheng, L.H. Tan, J.P. Chen and Y. Ting, "Biosorption performance of two brown marine algae for removal of chromium and cadmium", Journal of Dispersion Science and Technology, Vol. 25(5), pp. 679-686, 2005.
- [32] J. Wang and C. Chen, "Biosorbents for heavy metals removal and their future", Biotechnology Advances, Vol. 27, pp. 195–226, 2009.
- [33] S. Shanab, A. Essa and E. Shalaby, "Bioremoval capacity of three heavy metals by some micro algae species (Egyptian Isolates)", Plant Signaling & Behavior, Vol. 7(3), pp. 1–8, 2012.
- [34] R. Gupta, P. Ahuja, S. Khan, R.K. Saxena and H. Mohapatra, "Microbial biosorbents: Meeting challenges of heavy metal pollution in aqueous solutions", Current Science, Vol. 78 (8), pp. 967-973, 2000.
- [35] S. Dwivedi, "Bioremediation of heavy metals by algae: Current and future perspective", Journal of advanced laboratory research in biology, Vol. 3(3), 2013.
- [36] S. Sabat, R.V. Kavitha, S.L. Shantha, G, Nair, M. Ghosh, N. Chandroth and V.K. Murthy, "Biosorption: An Eco-Friendly Technique for the Removal of Heavy Metals." Indian Journal of Applied Research, Vol. 2(3), pp. 1-8, December 2012, ISSN - 2249-555X.
- [37] S. Radha, A. A. Fathima, S. Iyappan and M. Ramya, "Direct colony PCR for rapid identification of varied microalgae from freshwater environment", Journal of Applied Phycology, Vol. 25, pp. 609–613, 2013.
- [38] A. Godhe, D. M. Anderson and A. Holm, "PCR amplification of microalgal DNA for sequencing and species identification: studies on fixatives and algal growth stages", Harmful Algae, Vol. 1, pp. 375– 382, 2002.
- [39] S. Ghosh and N.G. Love, "Application of rbcL based molecular diversity analysis to algae in wastewater treatment plants", Bioresource Technology, Vol, 102, pp. 3619–3622, 2011.
- [40] C.B. Chikere, "Application of Molecular Microbiology Techniques in Bioremediation of Hydrocarbons and Other Pollutants", British Biotechnology Journal, Vol. 3(1), pp. 90-115, 2013.
- [41] M. Wan, J.N. Rosenberg, J. Farug, M.J. Betenbaugh and J, Xia, "An improved colony PCR procedure for genetic screening of Chlorella and related microalgae", Biotechnology Letters, Vol. 33, pp. 1615–1619, 2011.
- [42] F. Veglio and F. Beolchini, "Removal of metals by biosorption: A Review", Hydrometallurgy, Vol. 44: 301-316, 1997.
- [43] H.Wu, R. Hsen and L. Lin, "Identification of Chlorella spp. Isolates using ribosomal DNA sequencing. Botanical Bulletin of Academia Sinica", Vol. 42, pp. 115-121, 2001.
- [44] I. Ponnuswamy, S. Madhavan, Dr.Syed Shabudeen and U.S. Shoba, "Isolation and Identification of Green Microalgae for Carbon Sequestration & Waste Water Treatment by Using PCR Studies", International Journal of Engineering Science and Innovative Technology, Vol. 2(5), pp. 263-268, 2013.
 [45] G. Plaza, K. Ulfig, T.C. Hazen, R.L. Brigmon, "Use of Molecular
- [45] G. Plaza, K. Ulfig, T.C. Hazen, R.L. Brigmon, "Use of Molecular Techniques in Bioremediation", Acta Microbiologica Polonica, Vol. 50(3-4), pp. 205-218, 2001.

- [46] R.J. Watson and B. Blackwell, "Purification and Characterization of a common soil component which inhibits the polymerase chain reaction. Canandian Journal of Microbiology", Vol. 46, pp. 633-642, 2000.
- [47] X. Yang, P. Liu, Z. Hao, J. Shi, S. Zhang, "Characterization and identification of freshwater microalgal strains toward biofuel production", BioResources, Vol. 7(1), pp. 686–695, 2012.
- [48] J. Rajendhran, P. Gunasekaran P, "Microbial phylogeny and diversity: Small subunit ribosomal RNA sequence analysis and beyond", Microbiological Research, Vol. 166, pp. 99-110, 2011.
- [49] S.K. Ratha, R. Prasanna, V. Gupta, D.W. Dhar and A.K. Saxena, "Bioprospecting and indexing the microalgal diversity of different ecological habitats of India", World Journal of Microbiology and Biotechnology, Vol. 28, pp. 1657–1667, 2012.
- [50] S. Radha, A.A. Fathima, S. Iyappan and M. Ramya, "Direct colony PCR for rapid identification of varied microalgae from freshwater environment", Journal of Applied Phycology, Vol. 25, pp. 609–613, 2013.
- [51] Z. Liu, C. Liu, Y. Hou, S. Chen, D. Xiao, J. Zhang and F. Chen, "Isolation and Characterization of a Marine Microalga for Biofuel Production with Astaxanthin as a Co-Product", Energies, Vol. 6, pp. 2759-2772, 2013, doi:10.3390/en6062759
- [52] W. Ahilya, S. Anjali, "Application of *Padina tetrastromatica* for removal of heavy metal ions from wastewater", Journal of Environmental Research and Development, Vol. 7(2A), pp. 958-963, 2012.
- [53] N. Tamilselvan, K. Saurav and K. Kannabiran, "Biosorption of Cr (VI), Cr (III), Pb (II) and Cd (II) from Aqueous Solutions by Sargassum wightii and Caulerpa racemosa Algal Biomass", Journal of Oceanic and Coastal Sea Research, Vol. 11(1), pp. 52-58, 2012.
- [54] Brahmbhatt, N.H. Rinku, V. Patel and R.T. Jasrai, "Removal of cadmium, chromium and lead from filamentous alga of *Pithophora sp.* of industrial wastewater", International Journal of Environmental Sciences, Vol. 3(1), pp. 408-411, 2012.
- [55] B. Kızılkaya, G. Türker, R. Akgül and F. Doğan, "Comparative Study of Biosorption of Heavy Metals Using Living Green Algae *Scenedesmus quadricauda* and *Neochloris pseudoalveolaris*: Equilibrium and Kinetics", Journal of Dispersion Science and Technology, Vol. 33(3), pp. 410-419, 2012.
- [56] K. Chojnacka, A. Chojnacki and H. Gorecka, "Biosorption of Cr³⁺, Cd²⁺ and Cu²⁺ ions by blue-green algae *Spirulina* sp.: Kinetics, equilibrium and the mechanism of the process.", Chemosphere, Vol. 59, pp. 75–84, 2005.
- [57] Z. Aksu, T. Kutsal, "A comparative study for biosorption characteristics of heavy metal ions with *C. vulgaris*", Environmental Technology, Vol. 11, pp. 979-87, 1990.
- [58] Z. Aksu, D. Özer, H. I. Ekiz, T. Kutsal and A. Çaglar, "Investigation of biosorption of chromium (VI) on *C. crispata* in two-staged batch reactor", Environmental Technology, Vol. 17, pp. 215–220, 1996.
- [59] G.C. Donmez, Z. Aksu, A. Ozturk and T. Kutsal, "A comparative study on heavy metal biosorption characteristics of some algae", Process Biochemistry, Vol. 34, pp. 885–892, 1999.
- [60] E. Thirunavukkarasu and K. Palanivelu, "Biosorption of Cr(VI) from plating effluent using marine algal mass", Indian Journal of Biotechnology, Vol 6, pp 359-364, July 2007.
- [61] J. A. Priya, P. S. Rao, Y. Vijaya, A. S. Reddy and Professor A. Krishnaiah, "Biosorption of chromium(VI), nickel(II) and copper(II) ions from aqueous solutions using *Pithophora* algae", Toxicological & Environmental Chemistry, Vol. 89(3), pp. 421-442, 2007.

STATES	2008-09		2009-10		2010-11	
	Qty	Value	Qty	Value	Qty	Value
India	4073479	22633627	345580	10453620	4262207	22955675
Karnataka	4115	36475	6483	30856	8491	36851
Maharashtra	-	-	66	489	-	-
Odisha	4069364	2257152	3419031	10422275	4253716	22918824

TABLE 1: Production of Chromite , 2008-09 to 2010-11 (By States) (Qty in tonnes; value in Rs)

Source: Indian Minerals yearbook (2011) [8]

TABLE 2:	Uptake and Accumulation of Chromium by algal Species.

Biosorbent	Adsorption Capacity(mg/g)	References
Padina tetrastromatica	5.5	[52]
Sargassum weghtii (brown)	65.96	[53]
Pithophora sp.	4.9	[54]
Scenedesmus quadricanda	1.98-81.98	[55]
Spirulina sp	9.62	[56]
Chlorella vulgaris	23.6	[57]
Cladophora crispata	30.4	[58]
Chlorella vulagris	23.0	[59]
Scenedesmus obliquus	15.6	[59]
Synechocystis sp.	19.2	[59]
Padina boergesenli	49	[60]
Pithophora	434.10	[61]
Acid-treated Pithophora	666.21	[61]
Nizamuddina zanadinii	32.72	[22]
Stoechospermum marginatum	32.63	[22]
Cystoseira indica	43.38	[22]
Padina australis	37.82	[22]
Sargassum glauceseers	47.62	[22]
Spirogyra sp.	265	[26]
Desmococcus olivaceus	16	[11]
Scenedesmus quadricauda	12	[23]
Ceramium virgatum	26.5	[25]

FIGURE 1: Schematic showing molecular technique for environmental samples (soil, sediment and water) [42].

