

# Plant Growth Promoting Rhizobacteria and Humic Acid Improve Growth and Yield of Organically Grown Canola

Shakeel Ahmad<sup>1</sup>, Ihsanullah Daur<sup>1</sup>,  
Samir Gamil Al - Solaimani<sup>1</sup>, Sajid Mahmood<sup>1</sup>

<sup>1</sup>Department of Arid Land Agriculture,  
Faculty of Meteorology,  
Environment and Arid Land Agriculture,  
King Abdulaziz University,  
Jeddah, 80208, Kingdom of Saudi Arabia

Mohamed Hussein Madkour<sup>2</sup>  
<sup>2</sup>Department of Environmental Sciences,  
Faculty of Meteorology,  
Environment and Arid Land Agriculture,  
King Abdulaziz University, Jeddah 80208,  
Kingdom of Saudi Arabia.

**Abstract**— Plant bio-stimulant like plant growth promoting rhizobacteria (PGPR) and humic acid (HA) can boost crop production in organic farming due to their various features. This study was conducted to evaluate the potential of PGPR and HA for improving the effectiveness of cow and poultry manure which are very important nutrient sources. A field trial using canola as test crop was conducted according to split plot design in RCBD arrangement with four replications. Three levels of HA (HA<sub>0</sub>= No HA, HA<sub>10</sub>= 10 kg HA ha<sup>-1</sup> and HA<sub>20</sub>= 20 kg HA ha<sup>-1</sup>) as main plot factor and two levels of PGPR (P<sub>0</sub>= without PGPR and P<sub>1</sub>= with PGPR) as sub-plot factor were used. At the time of harvesting, data regarding growth and yield parameters such as plant height (cm), number of branches plant<sup>-1</sup>, number of pods plant<sup>-1</sup>, number of seeds pod<sup>-1</sup>, 1000 seed weight (g) and seed yield (t ha<sup>-1</sup>) was recorded and statistically analyzed using Statistix 8.1 computer software. It was observed that PGPR and HA improved all the growth and yield parameters. Significant interaction between PGPR and HA was also observed in their effect on 1000 seed weight and seed yield. It is concluded from the results that combined application of both PGPR and HA can improve growth and yield of organically grown canola more than sole application of each partner.

**Keywords**—PGPR; humic acid; cow manure; poultry manure; canola

## I. INTRODUCTION

Excessive and irrational use of chemical fertilizers and pesticides is putting the agricultural sustainability under great risk by deteriorating soil, water and environmental quality and health [1]. The ill effects of excessive use of fertilizers and pesticide include; pollution of ground water, negative impact on biodiversity, enhanced resistance of pests to chemicals, nutrient imbalance, soil salinization, impaired soil physical health resulting in erosion and soil degradation, reduced food quality due to entry of chemical residues, enhanced greenhouse gas emission and loss of natural habitat [2, 3]. The awareness about the negative impacts and issues generated by modern day agriculture gave birth to a sustainable farming approach called organic agriculture. Organic farming is the agricultural management system that excludes the use of synthetic chemicals (fertilizers and pesticides) and employs such nutritional (crop rotation, biological nitrogen fixation, cover cropping, use of animal manures, non-synthetic and unprocessed minerals), plant protection (biopesticides) and

cultural practices which not only improve crop productivity and produce quality but also ensure sustainability by enhancing soil quality and health [4, 5].

Management of fertility for optimum plant nutrition is big challenge in organic agriculture. Animal manures like cow manure (CM) and poultry manure (PM) are considered as important nutrient sources in organic farming. But there are some issues which reduce the efficiency of manures as good nutrient sources. Poultry manures have been reported to cause burning effects on crops due to its high nutrient and salt contents [6]. Similarly, poor nutrient use efficiency due to slow nutrient release which does not synchronize with plant need is another important issue [7, 8]. Prevailing scenario prompted us to devise some cost effective and sustainable approaches to enhance the effectiveness of manures. We thought that problem of burning effect of PM can be overcome by mixing it with cow manure. Similarly, after extensive review of literature we found that plant bio-stimulants like plant growth promoting rhizobacteria (PGPR) and humic acids (HA) can work as bio-activators to enhance the efficacy of manures due to their various promising features.

PGPR is a diverse group of free or associatively living soil bacteria that inhabit in the vicinity of plant roots and exert positive effects on plant growth, development and yield by the production of plant growth regulators, biological nitrogen fixation, solubilization of nutrients, mineralization of nutrient from organic matter through decomposition and enzymatic release and better nutrient uptake due to improved root growth [9-11]. Similarly, HA are components of humic substances (HS) that are end products of microbial decomposition and chemical degradation of dead biota [12]. HA can promote plant growth through improvement in root architecture and morphology, enhancement of nutrient availability, uptake and translocation, promotion of plant physiology and development due to their hormone-like activities and promotion of soil biological activity [13-18].

Keeping in view the characteristics of PGPR and HA we hypothesized that they can enhance the efficacy of manures through the activation of natural processes like nutrient release and uptake by various mechanisms described above. Extensive root system created by improved root growth and

architecture by PGPR and HA can also enhance the effectiveness of manures by increasing the uptake of nutrients by exploring more and more soil volume. Stimulation of biological activity by HA can further enhance nutrient cycling through the action of microorganisms. Therefore to evaluate the potential of PGPR and HA for increasing effectiveness of manures we planned a field experiment using canola crop. In this experiment we used mixture of manures (CM and PM) prepared by blending the CM and PM in 3:1 to overcome the issue of burning effect of poultry manure.

## II. MATERIALS AND METHODS

### A. Experimental site and soil characteristic

The experiment was conducted at Agricultural Research Station of King Abdulaziz University, Hada Al-Sham (21o 48' 3" N, 39o 43' 25"E). Analysis of soil was carried out according to procedure given in Soil and Plant Analysis Laboratory Manual by Ryan and his colleagues [19]. According to results of analysis, soil was loamy sand in texture having pH (7.68), electrical conductivity ( $3.24 \text{ dS m}^{-1}$ ) and organic matter (0.65%). Nutrient analysis of soil revealed that it contains 0.054% total N,  $6.4 \text{ mg kg}^{-1}$  available P,  $355 \text{ mg kg}^{-1}$  extractable K and 54, 17, 2 and  $5 \text{ mg kg}^{-1}$  Fe, Zn, Mn and Cu respectively.

### B. Collection of PGPR strain and seed inoculation

Pre-isolated strain of PGPR having plant growth promoting traits (IAA production, phosphate solubilization, ACC-deaminase activity and siderophore production) characterized by Ahmad and his co-workers [20] was used in this study. Broth culture of PGPR strain for seed inoculation was prepared in tryptic soy broth (TSB) medium according to Atlas [21]. Seed inoculation was done according to Baig and his co-workers [22]. First of all canola seeds were surface sterilized with 95% ethanol by dipping the seeds in ethanol for short time. After taking out the seeds from ethanol, they were washed with sterilized water and were immersed in 0.2%  $\text{HgCl}_2$  solution for 3 minutes and washed again with sterilized water. Then surface sterilized seeds were inoculated with the broth mixed with 10% sugar solution, peat and clay (Kaolin) mixture (peat to clay ratio, 1:1 w/w). The seeds were shaken well until a fine coating appeared on seeds. Control seeds were treated with sterilized peat plus clay mixed with sterilized broth medium (without bacterial cells) and sugar solution. Inoculated seeds were placed overnight for drying under laboratory conditions at  $26^\circ\text{C}$ .

### C. Field trial and treatment plan

To evaluate the combined effect of PGPR and HA for improving growth and yield of organically grown canola, a field trial was conducted according to split plot design in RCBD arrangement with four replications. Three levels of HA ( $\text{HA}_0 = \text{No HA}$ ,  $\text{HA}_{10} = 10 \text{ kg HA ha}^{-1}$  and  $\text{HA}_{20} = 20 \text{ kg HA ha}^{-1}$ ) as main plot factor and two levels of PGPR ( $\text{P}_0 = \text{without PGPR}$  and  $\text{P}_1 = \text{with PGPR}$ ) as sub-plot factor were used. First of all field was ploughed well with rotary plough. After that, 6 month old naturally composted cow manure (CM) and poultry manure (PM) were obtained from Agricultural and Dairy Research Station of King Abdulaziz University, Hada Al-Sham. Then manures were mixed well in 3:1 (CM: PM) to get a fine blend. After getting a fine manure blend it was distributed and mixed well in the field @  $15 \text{ t ha}^{-1}$  and main and sub plots were marked. Then different rates of humic acid were applied in the soil according to layout and

mixed well in the soil. Finally, inoculated seeds were sown by keeping row to row and plant to plant distances of 40 cm and 30 cm respectively. Recommended cultural and agronomic practices were adopted for irrigation, weeding, plant protection and harvesting.

### D. Data collection and statistical analysis

At the time of harvesting, data regarding growth and yield parameters such as plant height (cm), number of branches plant<sup>-1</sup>, number of pods plant<sup>-1</sup>, number of seeds pod<sup>-1</sup>, 1000 seed weight (g) and seed yield ( $\text{t ha}^{-1}$ ) were recorded to evaluate the effect of the treatments on crop growth and yield. All data was recorded according to standard procedures. Before harvesting, 10 random guarded plants in each sub-sub plot were tagged to record data regarding plant height (cm), number of branches plant<sup>-1</sup>, number of pods plant<sup>-1</sup> and number of seeds pod<sup>-1</sup>. For seed yield an area of  $1 \text{ m}^2$  in the center of each sub-sub plot (to avoid marginal effect) was harvested. Data recorded for each trait was separately exposed to the analysis of variance (ANOVA) to check significance among treatments using Statistix 8.1 computer software and means were then compared using least significant difference (LSD) test according to Steel and Torrie [23].

## III. RESULTS

The analyzed data regarding the combined effect of HA and PGPR on growth and yield attributes of organically grown canola is given in Table 1. Results revealed that all the studied parameters were significantly improved by humic acid and PGPR application. However interaction of HA and PGPR was found significant only for 1000 seed weight and seed yield.

Mean comparison between different rates of HA (0, 10 and  $20 \text{ kg HA ha}^{-1}$ ) for their effect on plant height showed that application of HA @  $20 \text{ kg ha}^{-1}$  produced maximum plant height (102 cm) that was significantly ( $p \leq 0.05$ ) higher compared to plots that received 0 and  $10 \text{ kg HA ha}^{-1}$ . Similarly, application of PGPR significantly induced more plant height (99 cm) than uninoculated treatment (96 cm). Number of branches per plant also showed similar trend. Maximum number of branches (26) was produced with  $20 \text{ kg HA ha}^{-1}$ , while 0 and  $10 \text{ kg HA ha}^{-1}$  produced 21 and 23 branches respectively. PGPR application produced more number of branches (25) than no PGPR application (22).

Number of pods plant<sup>-1</sup> showed somewhat different trend where  $20 \text{ kg HA ha}^{-1}$  with the production of 151 pods was statistically superior to both 0 and  $10 \text{ kg HA ha}^{-1}$ . Here,  $10 \text{ kg HA ha}^{-1}$  by producing 123 pods was also significantly higher than  $0 \text{ kg HA ha}^{-1}$  which produced 108 number of pods. Likewise, PGPR application also proved statistically better than no PGPR. PGPR application produced 134 pods while uninoculated control produced only 121 pods per plant. Regarding number seeds pod<sup>-1</sup>, statistically higher numbers of seeds (27) were observed for  $20 \text{ kg HA ha}^{-1}$  followed by 22 and 19 seed pod<sup>-1</sup> for 10 and  $0 \text{ kg HA ha}^{-1}$  respectively. PGPR inoculation also showed promising effects for improving number of seeds pod<sup>-1</sup> and produced significantly higher (24) seeds pod<sup>-1</sup> than the treatment without PGPR that produced 21 seeds pod<sup>-1</sup>.

Data regarding the effect of HA and PGPR on 1000 seed weight of canola indicated that both humic acid and PGPR have interactive effect to improve this parameter. Interaction effect of PGPR and HA for 1000 seed weight presented in

Table 2 clearly indicated that maximum value of 1000 seed weight (2.45 g) was observed with 20 kg HA ha<sup>-1</sup> at both levels of PGPR. Minimum value of 1000 seed weight (2.36 g)

Table 1: Effect of humic acid and PGPR on growth and yield parameters of canola

Treatments	Plant Height (cm)	No. of branches plant <sup>-1</sup>	No. of pods plant <sup>-1</sup>	No. of seeds pod <sup>-1</sup>	1000-seed weight (g)	Seed yield (t/ha)
Humic acid (kg/ha)						
HA <sub>0</sub> (0 kg)	94 <sup>b</sup>	21 <sup>b</sup>	108 <sup>c</sup>	19 <sup>b</sup>	2.38 <sup>b</sup>	1.54 <sup>c</sup>
HA <sub>10</sub> (10 kg)	97 <sup>b</sup>	23 <sup>b</sup>	123 <sup>b</sup>	22 <sup>b</sup>	2.43 <sup>a</sup>	1.86 <sup>b</sup>
HA <sub>20</sub> (20 kg)	102 <sup>a</sup>	26 <sup>a</sup>	151 <sup>a</sup>	27 <sup>a</sup>	2.45 <sup>a</sup>	2.40 <sup>a</sup>
LSD	4	2.33	2	3.9	0.04	0.11
PGPR						
P <sub>0</sub> (No PGPR)	96 <sup>b</sup>	22 <sup>b</sup>	121 <sup>b</sup>	21 <sup>b</sup>	2.41 <sup>b</sup>	1.70 <sup>b</sup>
P <sub>1</sub> (PGPR)	99 <sup>a</sup>	25 <sup>a</sup>	134 <sup>a</sup>	24 <sup>a</sup>	2.43 <sup>a</sup>	2.16 <sup>a</sup>
LSD	1.9	2.46	5	2.14	0.01	0.16
Significance						
Humic acid	**	**	**	**	**	**
PGPR	**	*	**	*	**	**
HA * PGPR	ns	ns	ns	ns	**	*

Means followed by the same letter (s) in each column and treatment showed no significant difference.

\*, \*\* indicate significant differences at 0.05, 0.01 probability levels respectively while ns indicate non-significant difference.

Table 2: Interaction effect of PGPR and humic acid

Humic acid (HA)	PGPR (P)	1000-seed weight (g)	Seed Yield (t/ha)
HA <sub>0</sub>	P <sub>0</sub>	2.36 <sup>c</sup>	1.37 <sup>e</sup>
	P <sub>1</sub>	2.40 <sup>b</sup>	1.70 <sup>d</sup>
HA <sub>10</sub>	P <sub>0</sub>	2.42 <sup>b</sup>	1.72 <sup>cd</sup>
	P <sub>1</sub>	2.44 <sup>ab</sup>	2.00 <sup>bc</sup>
HA <sub>20</sub>	P <sub>0</sub>	2.45 <sup>a</sup>	2.02 <sup>b</sup>
	P <sub>1</sub>	2.45 <sup>a</sup>	2.79 <sup>a</sup>
	LSD	0.02	0.28

was recorded with the combination of 0 kg HA ha<sup>-1</sup> and no PGPR application.

As seed yield is closely related to yield components, so like yield components (pods plant<sup>-1</sup>, number of seeds pod<sup>-1</sup>, and 1000 seed weight) HA and PGPR application also showed significant effect on this parameter. For seed yield, significant ( $p \leq 0.05$ ) interaction between HA and PGPR was observed (Table 2). The results presented in table clearly demonstrated that application of PGPR at each level of humic acid has produced more seed yield than without PGPR at the same level of humic acid. Maximum seed yield (2.79 t ha<sup>-1</sup>) was achieved with the combined application of PGPR and HA @ 20 kg ha<sup>-1</sup>, which is significantly higher than all other interactions. Minimum seed yield (1.37 t ha<sup>-1</sup>) was observed where no PGPR and HA was applied.

#### IV. DISCUSSION

It was confirmed from the results that combined application of PGPR and HA significantly improved growth and yield attributes of canola under field conditions. Previously, PGPR have been reported to enhance plant growth and yield by the production of PGRs (IAA, gibberellins, and cytokinins etc.) and improved nutrient availability and uptake by nutrient solubilization, nutrient cycling and enhanced root growth [24, 25, 9]. Possession of plant growth promoting activities by the strain used in this study already confirmed by Ahmad and his co-workers [20] further strengthens our results.



The impact of HA growth and yield parameters may be reasoned to its hormone-like activity and role in plant metabolism [13, 26]. Auxin-like activity of HA has also been reported earlier by some researchers [27, 16]. It is an established fact that PGRs accelerate the vital developmental processes of cell elongation, cell division which cause improvement in plant growth [28]. Moreover, both HA and PGPR have been reported to indirectly promote plant growth parameters by enhancing nutrient availability and uptake especially N which has significant role in the promotion of vegetative growth [29].

Different researchers have reported the effects of HA and PGPRs on various physiological and metabolic processes, nutrients uptake and translocation and accumulation of photosynthates. For example, El-Nemr and his co-workers [30] conducted a field trial on cucumber in Egypt for two seasons and noted that humic acid significantly increased number of flowers and fruit per plant, fruit set, mean fruit weight, fruit length and diameter, and yield per plant. Likewise, Yildirim [31] observed improvement in growth and yield parameters of tomato by the accumulation of total soluble solids and ascorbic acid contents with humic acid application. Similarly, Karakurt and his co-workers [32] recorded enhancement in total soluble sugars, reducing sugars, and chlorophyll b content of pepper with parallel improvement in growth and yield by humic acid application. Findings of our work regarding improvement in growth and yield parameters by PGPR are also in line with other researchers [33-35] who have reported improvement in growth and yield of crops by PGPR.

#### V. CONCLUSION

The results of this study are very promising and inspiring. The improvement of canola growth and yield by the combined application of PGPR and in this region is of great value. Because, canola can be successfully grown in dry land or under irrigation even with saline soil conditions. Application of this approach at large scale can be helpful to boost canola production to overcome the problem of fodder shortage.

#### ACKNOWLEDGMENT

Authors are thankful to Deanship of Scientific Research and Chairman, Department of Arid Land Agriculture, King Abdulaziz University for providing facilities for experimentation.

#### REFERENCES

1. Tuomisto, H. L., Hodge, I. D., Riordan, P., & Macdonald, D. W. (2012). Does organic farming reduce environmental impacts?—A meta-analysis of European research. *Journal of environmental management*, 112, 309-320.
2. Azizullah, A., Khattak, M. N. K., Richter, P., & Häder, D. P. (2011). Water pollution in Pakistan and its impact on public health—a review. *Environment International*, 37(2), 479-497.
3. Conway, G. R., & Pretty, J. N. (2013). *Unwelcome harvest: agriculture and pollution*. Routledge.
4. Diacono, M., & Montemurro, F. (2010). Long-term effects of organic amendments on soil fertility. A review. *Agronomy for sustainable development*, 30(2), 401-422.
5. Paungfoo-Lonhienne, C., Visser, J., Lonhienne, T. G., & Schmidt, S. (2012). Past, present and future of organic nutrients. *Plant and Soil*, 359(1-2), 1-18.
6. Ellis, L., S. Love, A. Moore and M.E. de Haro-Marti. 2013. Composting and using backyard poultry waste in home garden. *CIS1194*. University of Idaho Extension.

7. Aulakh, M. S. (2010). Integrated nutrient management for sustainable crop production, improving crop quality and soil health, and minimizing environmental pollution. In *19th world congress of soil science, soil solutions for a changing world, Brisbane* (pp. 1-6).
8. Leconte, M. C., Mazzarino, M. J., Satti, P., & Crego, M. P. (2011). Nitrogen and phosphorus release from poultry manure composts: the role of carbonaceous bulking agents and compost particle sizes. *Biology and Fertility of Soils*, 47(8), 897-906.
9. Bhattacharyya, P. N., & Jha, D. K. (2012). Plant growth-promoting rhizobacteria (PGPR): emergence in agriculture. *World Journal of Microbiology and Biotechnology*, 28(4), 1327-1350.
10. Ahemad, M., & Kibret, M. (2014). Mechanisms and applications of plant growth promoting rhizobacteria: Current perspective. *Journal of King Saud University-Science*, 26(1), 1-20.
11. Miransari, M. (2014). Plant growth promoting rhizobacteria. *Journal of Plant Nutrition*, 37(14), 2227-2235.
12. Schiavon M, Pizzeghello D, Muscolo A, Vaccoro S, Francioso O, Nardi S (2010) High molecular size humic substances enhance phytylpropanoid metabolism in maize (*Zea mays* L.). *J Chem Ecol* 36:662–669.
13. Nardi S, Carletti P, Pizzeghello D, Muscolo A (2009) Biological activities of humic substances. In: Senesi N, Xing B, Huang PM (ed) *Biophysico-chemical processes involving natural nonliving organic matter in environmental systems*. Vol 2, part 1: fundamentals and impact of mineral-organic biota interactions on the formation, transformation, turnover, and storage of natural nonliving organic matter (NOM). Wiley, Hoboken, pp 305–339
14. Ferrara, G. and Brunetti, G. (2010) Effects of the Times of Application of a Soil Humic Acid on Berry Quality of Table Grape (*Vitis vinifera* L.) c.v Italia. *Spanish Journal of Agricultural Research*, 8, 817-822.
15. Trevisan, S., Francioso, O., Quaggiotti, S., & Nardi, S. (2010). Humic substances biological activity at the plant-soil interface: from environmental aspects to molecular factors. *Plant signaling & behavior*, 5(6), 635-643.
16. Jindo K, Martim SA, Navarro EC et al (2012) Root growth promotion by humic acids from composted and noncomposted urban organic wastes. *Plant Soil* 353:209–220
17. Puglisi E, Pascasio S, Suciù N, Cattani I, Fait G, Spaccini R, Crecchio C, Piccolo A, Trevisan M (2013) Rhizosphere microbial diversity as influenced by humic substance amendments and chemical composition of rhizodeposits. *J Geochem Expl* 129:82–94.
18. Canellas, L. P., & Olivares, F. L. (2014). Physiological responses to humic substances as plant growth promoter. *Chemical and Biological Technologies in Agriculture*, 1(1), 1-11.
19. Ryan, J., G. Estefan and A. Rashid. 2001. *Soil and plant analysis: Laboratory manual. International Centre for Agricultural Research in the Dry Areas*. Aleppo, Syria.
20. Ahmad, S., Daur, I., Al-Solaimani, SG., Mahmood, S., Madkour, MH. 2016. Exploiting the Potential of Indigenous Plant Growth Promoting Rhizobacteria (PGPR) for Improving Growth and Yield Attributes of Canola in Western Saudi Arabia. *Int. J. Agric. Inno. Res.* 4: 955-959.
21. Atlas, R. M. (2010). *Handbook of microbiological media*. CRC press.
22. Baig, K. S., Arshad, M., Shaharouna, B., Khalid, A., & Ahmed, I. (2012). Comparative effectiveness of *Bacillus* spp. possessing either dual or single growth-promoting traits for improving phosphorus uptake, growth and yield of wheat (*Triticum aestivum* L.). *Annals of microbiology*, 62(3), 1109-1119.
23. Steel, R. D., Torrie, J. H., & Dickey, T. A. (1997). Principles and practice of statistics: a biomedical approach. *New York, USA, McGrawHill*, 297-299.
24. Barea, J. M., Azcón, R., & Azcón-Aguilar, C. (2005). Interactions between mycorrhizal fungi and bacteria to improve plant nutrient cycling and soil structure. In *Microorganisms in soils: roles in genesis and functions* (pp. 195-212). Springer Berlin Heidelberg.
25. Singh, J. S., Pandey, V. C., & Singh, D. P. (2011). Efficient soil microorganisms: a new dimension for sustainable agriculture and environmental development. *Agriculture, Ecosystems & Environment*, 140(3), 339-353.
26. Berbara, R.L.L. and A.C. García. 2014. Humic substances and plant defense metabolism. In: Ahmad, P. and M.R. Wani (eds.) *Physiological mechanisms and adaptation strategies in plants under changing environment*. Springer Science+Business Media, New York, pp. 297-319.
27. Canellas, L.P., D.J. Dantas, N.O. Aguiar, L.E.P. Peres, A. Zsögön, F.L. Olivares, L.B. Dobbss, A.R. Façanha, A. Nebbioso and A. Piccolo. 2011. Probing the hormonal activity of fractionated molecular humic components in tomato auxin mutants. *Ann. Appl. Biol.*, 159(2):202-211.

28. Campanoni, P. and P. Nick. 2005. Auxin-dependent cell division and cell elongation. 1-Naphthaleneacetic acid and 2, 4-dichlorophenoxyacetic acid activate different pathways. *Plant Physiol.*, 137(3): 939-948.
29. Metay, A., J. Magnier, N. Guilpart and A. Christophe. 2015. Nitrogen supply controls vegetative growth, biomass and nitrogen allocation for grapevine (cv. Shiraz) grown in pots. *Funct. Plant Biol.*, 42(1): 105-114.
30. El-Nemr, M.A., M. El-Desuki, A.M. El-Bassiony and Z.F. Fawzy. 2012. Response of growth and yield of cucumber plants (*Cucumis sativus*L.) to different foliar applications of humic acid and bio-stimulators. *Aust. J. Basic Appl. Sci.*, 6:630-637.
31. Yildirim, E. 2007. Foliar and soil fertilization of humic acid affect productivity and quality of tomato. *Acta. Agric. Scand. Sect. B. Soil Plant Sci.*, 57(2): 182-186.
32. Karakurt, Y., H. Unlu, H. Unlu and H. Padem. 2009. The influence of foliar and soil fertilization of humic acid on yield and quality of pepper. *Acta. Agric. Scand. Sect. B. Soil Plant Sci.*, 59(3): 233-237.
33. Verma, V.C., S.K. Singh and S. Prakash. 2011. Bio-control and plant growth promotion potential of siderophore producing endophytic *Streptomyces* from *Azadirachta indica* A. Juss. *J. Basic Microbiol.*, 51(5): 550-556.
34. Shahzad, S.M., M.S. Arif, M. Riaz, Z. Iqbal and M. Ashraf. 2013. PGPR with varied ACC-deaminase activity induced different growth and yield response in maize (*Zea mays* L.) under fertilized conditions. *Eur. J. Soil Biol.*, 57: 27-34.
35. Zahid, M., M.K. Abbasi, S. Hameed and N. Rahim. 2015. Isolation and identification of indigenous plant growth promoting rhizobacteria from Himalayan region of Kashmir and their effect on improving growth and nutrient contents of maize (*Zea mays* L.). *Front. Microbiol.*, 6.